Stem cell fate in cancer growth, progression and therapy resistance

Nikki K. Lytle^{1,2,3}, Alison G. Barber^{1,2,3} and Tannishtha Reya^{1,2,3}*

Abstract | Although we have come a long way in our understanding of the signals that drive cancer growth, and how these signals can be targeted, effective control of this disease remains a key scientific and medical challenge. The therapy resistance and relapse that are commonly seen are driven in large part by the inherent heterogeneity within cancers that allows drugs to effectively eliminate some, but not all, malignant cells. Here, we focus on the fundamental drivers of this heterogeneity by examining emerging evidence that shows that these traits are often controlled by the disruption of normal cell fate and aberrant adoption of stem cell signals. We discuss how undifferentiated cells are preferentially primed for transformation and often serve as the cell of origin for cancers. We also consider evidence showing that activation of stem cell programmes in cancers can lead to progression, therapy resistance and metastatic growth and that targeting these attributes may enable better control over a difficult disease.

Stem cell

A cell that has the ability to perpetuate itself through self-renewal and to generate differentiated cells. Stem cells are relatively rare among other cell types and can be more quiescent and resistant to toxins and chemicals as well as display enhanced DNA repair.

Over the past several decades, cancer has largely been treated as a disease of aberrant proliferation and survival, and the therapies most commonly used today — radiation and chemotherapy — mainly target these properties. Despite the successes of cytotoxic therapies, which include cures achieved in childhood acute lymphoblastic leukaemia (ALL)¹ and lymphoma², it is also clear that we are reaching the limits of how effective these approaches can be, at least in their current form. Thus, it has become important to examine aspects of oncogenesis beyond aberrant survival and proliferation.

One critical aspect of the changes that occur as benign lesions transition to malignant ones is the progressive acquisition of the undifferentiated state. Benign lesions are often more differentiated, while malignancies are more undifferentiated, suggesting a reversal of the differentiation signals put in place during development. As many of the signals that drive the undifferentiated state are also key to conferring a stem cell fate, it is perhaps not surprising that many cancers show an acute dependence on these signals to maintain their more aggressive state.

In addition, stem cell signals are also integrally linked to cancer initiation, propagation and therapy resistance. While driver mutations are key to initiating oncogenesis, the cells in which these mutations occur are of equal importance; thus, mutations that cannot transform differentiated cells can transform undifferentiated ones^{3–6}, suggesting that the stem or progenitor cell state provides a more permissive context for transformation. Even after cancer establishment, perpetuation of a stem cell state in some cells creates cancer stem cells (CSCs), 'driver cells'

that are preferentially aggressive and pose a high risk of therapy resistance and disease relapse⁷. Thus, understanding and targeting the signals critical to sustaining these cells are essential to improving outcomes. Here, we focus on how regulation of stem cell fate can not only influence cancer initiation but also serve as a driver event for disease progression, therapy resistance and metastatic growth.

Stem cell states in cancer initiation

Transcriptional context and the cell of origin. Key studies have shown that subsets of cells with stem and progenitor characteristics in normal tissues are particularly susceptible to oncogenic transformation. Beginning with work in haematological malignancies, in which chronic myeloid leukaemia (CML) arose only when the BCR-ABL mutation occurred in stem cells^{8,9}, this paradigm has now been extended to other haematological 10,11 and solid cancers3,12. Defining the cell of origin can be critical for understanding both the environments that are permissive for transformation and the signals required for transformation. BCR-ABL provides an interesting example of an oncogene that produces different outcomes depending on the cell in which it is expressed. While BCR-ABL rapidly triggered CML when introduced into stem cells, it triggered B cell ALL (B-ALL) when expressed in progenitor cells13. Interestingly, this difference in cell of origin is closely coupled to differential signal dependencies: while loss of β -catenin blocked CML propagation, it did not impact B-ALL to the same extent¹³. Thus, the cell of origin can define both the nature of the cancer and its dependencies.

- Departments of Pharmacology and Medicine, San Diego School of Medicine, University of California, La Jolla, CA, USA.
- ²Sanford Consortium for Regenerative Medicine, San Diego School of Medicine, University of California, La Jolla. CA. USA.
- ³Moores Cancer Center, San Diego School of Medicine, University of California, La Jolla, CA, USA.
- *e-mail: treya@ucsd.edu https://doi.org/10.1038/ s41568-018-0056-x

Stem cell signals

Also called stem cell programmes, these are signals or gene expression programmes that are often associated with the undifferentiated state in embryonic and adult stem cells. Many stem cell programmes or signalling pathways are reactivated in oncogenesis.

Cancer stem cells

(CSCs). Cells with enriched functional capacity to drive tumour growth and recreate its heterogeneity. CSCs generally share many of the defining characteristics of normal stem cells, including increased drug resistance and DNA repair.

The link between the cell of origin and tumour types holds true across some solid cancers. For example, expression of KRAS in the context of p53 loss triggered squamous cell carcinoma when targeted to either interfollicular epidermis cells or hair follicle cells¹⁴. Despite both cell types giving rise to squamous cell carcinoma. interfollicular epidermis-derived tumours were largely epithelial in nature and had limited metastatic potential, whereas hair follicle-derived tumours contained a mixture of both epithelial and mesenchymal tumour cells and were associated with a higher incidence of metastasis¹⁴. Similarly, the cell of origin in glioblastoma can dictate sensitivity to transformation and the type of tumour formed. Concurrent inactivation of Trp53, Nf1 and Pten in neural stem cells, neural progenitor cells or oligodendrocyte progenitors triggered the development of unique subtypes of glioblastoma with distinct gene expression profiles that were predictive of differential molecular dependencies^{15,16}. These studies suggest that the transcriptional context of the cell of origin can be selectively permissive for specific tumour types and can be at least as strong a determinant of tumorigenesis as the driver mutations themselves (FIG. 1a).

By contrast, activation of Hedgehog signalling via genetic deletion of its receptor, patched homologue 1 (PTC1, which is encoded by Ptch1), in either neural stem cells or granule neural precursors leads to development of aggressive medulloblastomas with similar molecular profiles¹⁷. This suggests that in some cases certain driver mutations, rather than the cell of origin, define the tumour profile by leading to a convergence of cell states^{17,18} (FIG. 1b). However, it remains unclear which tumours are predominantly determined by the cell of origin versus by the relevant mutations. It is possible that certain mutations are powerful enough in terms of defining cell fate that they can override the transcriptional context of the cell of origin; for example, in the cases above, mutations in the Hedgehog pathway could have a dominant impact on fate because they can control

stem cell programmes (FIG. 1b). Given the impact of these early tumorigenic events in determining tumour evolution, it may be important to better understand the factors that control tumour cell fate.

Epigenetic mechanisms and the cell of origin. In addition to the transcriptional context dictating susceptibility to transformation, epigenetic states may also direct tumour-initiating capacity and mutational state. Recent studies have shown that changes in DNA¹⁹ or histone²⁰ methylation patterns can override oncogene-induced senescence programmes. Moreover, transformed cells that escape senescence have increased DNA methylation, leading to inactivation at promoters of differentiation-associated genes¹⁹. This suggests that the epigenetic landscape is a critical determinant of both transformation susceptibility and the acquisition of a stem or progenitor phenotype.

Work in zebrafish models has shown that there is an early permissive epigenetic signature within tumourinitiating cells in melanoma²¹. In a field of melanocytes expressing driver mutations, only cells harbouring an epigenetic profile that enabled SRY-box 10 (Sox10)driven expression of the fetal oncogene crestin were sensitive to transformation²¹. Furthermore, chromatin accessibility in melanocytes substantially overlapped with mutational density in human melanoma samples, suggesting that the combination of mutations that drive tumorigenesis mirrors the permissive epigenetic landscape of the cells within the tissue of origin²². Consistent with this, the epigenetic landscape of normal cells and the mutational status of tumours from the same tissue were also congruent in liver cancer, multiple myeloma, colorectal cancer, oesophageal cancer, glioblastoma, lung adenocarcinoma and lung squamous cell carcinoma²². Highlighting the importance of the tumourinitiating cell in defining the molecular profile of the tumour, a survey of over 10,000 tumour samples across

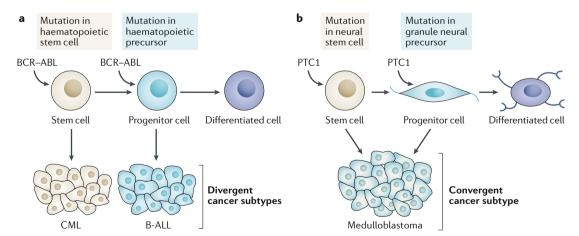


Fig. 1 | Impact of the cell of origin on cancer development. a | Oncogenic mutation can drive distinct cancer subtypes depending on the epigenetic and transcriptomic profile of the cell of origin. For example, in haematological malignancies, when BCR-ABL is introduced into stem cells, it results in chronic myeloid leukaemia (CML); however, when this same mutation is introduced into progenitor cells, it results in B cell acute lymphoblastic leukaemia (B-ALL). b | Alternatively, oncogenic mutation in distinct cells of origin can lead to a convergence of cell states that results in the same cancer subtype. For example, in medulloblastoma, deletion of patched homologue 1 (PTC1) in either neural stem cells or granule neural precursors leads to the development of aggressive medulloblastoma.

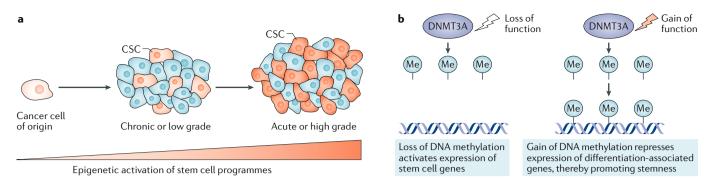


Fig. 2 | Epigenetic regulation of the stem cell state in cancer. a | During normal development, stem cell programmes are extinguished during differentiation; in cancers, such as myeloid leukaemia, epigenetic reactivation of stem cell programmes can promote propagation and progression to an aggressive state. The activation of these programmes in a subpopulation (cancer stem cells (CSCs), shown in orange) is associated with chronic myeloid leukaemia (CML), a low-grade disease, while widespread activation of these programmes — illustrated by the expanded pool of CSCs in the figure — is associated with blast crisis CML, an aggressive, high-grade disease. b | Epigenetic regulation of stem cell programmes may also be mediated through modification of DNA. For example, mutation of DNA (cytosine-5)-methyltransferase 3A (DNMT3A) can promote the stem cell state through either loss of function mutations (which can lead to hypomethylation and activation of genes that promote the stem cell state; shown on the left) or gain of function mutations (which can lead to hypermethylation and silencing of genes associated with differentiation; shown on the right). Me, methylation.

cancers revealed that the methylome, transcriptome and proteome all cluster by the tissue of origin²³.

Importantly, the epigenetic alterations that precede transformation may act as the functional equivalent of a driver mutation. Bronchial epithelial cells chronically exposed to cigarette smoke display altered methylation patterns that lead to aberrant KRAS, WNT and epidermal growth factor receptor (EGFR) signalling²⁴. The altered epigenetic state sensitized the cells to transformation with just one mutation instead of the three normally required²⁴. Thus a deeper understanding of how epigenetic mechanisms contribute to the acquisition or maintenance of a stem cell phenotype is critical for developing strategies to disable the early permissive states for effective early intervention or prevention strategies.

Stem cell states in tumour propagation

Genetic and epigenetic control of cell fate in cancer. Beyond their role in establishing the cell of origin and initiating oncogenesis, programmes that control cell fate are critical for cancer propagation via both genetic and epigenetic mechanisms. Multiple stem cell signals such as WNT or NOTCH or those of the Hedgehog pathway are activated in various cancers through mutations. For example, loss of adenomatous polyposis coli (APC) in colon cancer activates the WNT pathway²⁵, activating mutations in smoothened homologue (SMO) or gliomaassociated oncogene (GLI1), or loss of PTC1 trigger aberrant Hedgehog signalling in medulloblastoma²⁶ and basal cell carcinoma²⁷, and NOTCH mutations are prevalent in T cell ALL (T-ALL)28; in each of these contexts, the signals serve as driver mutations, highlighting the powerful influence of stem cell signals in promoting oncogenic growth.

While in some cancers genes encoding members of stem cell signalling pathways are recurrently mutated, in other cancers these same genes are often activated epigenetically (FIG. 2a). For example, *NOTCH1* is epigenetically

activated in breast cancers and pancreatic cancer^{29,30}, as is WNT signalling in leukaemias31, and targeting these factors therapeutically using monoclonal antibodies against the NOTCH ligand delta-like protein 4 (DLL4) or antagonists of CREB-binding protein (CBP) and β-catenin, respectively, is currently being tested in clinical trials^{32,33}. More recently, defined stem cell signals, such as the RNA-binding protein Musashi homologue (MSI), have also been shown to be both genetically and epigenetically modified in cancers; for example, blast crisis CML can harbour translocations in MSI2 (REF.34), but MSI2 can also be epigenetically activated in the absence of mutations^{34–36}. The discussion below focuses on how epigenetic mechanisms can influence expression and activation of stem cell signalling pathways to support cancer propagation.

DNA methylation can also influence the acquisition of the stem cell state in cancer. Alterations in DNA methylation may occur early in tumour development: inactivating mutations in DNMT3A (which encodes DNA (cytosine-5)-methyltransferase 3A) lead to altered methylation and leukaemia onset^{37,38}, and Dnmt3A deletions can trigger a spectrum of haematological malignancies in mouse models³⁹⁻⁴¹ (FIG. 2b). At a molecular level, mutant DNMT3A leads to decreased DNA methylation⁴², which may confer stem cell fate by activating stem cell genes such as those in the HOXB cluster43 and leaving pro-differentiation genes hypermethylated³⁷. Promoter hypomethylation may also be a mechanism by which other key stem cell genes are reactivated in high-grade cancer; for example, hypomethylation of the MSI1 locus is linked to high expression in triple-negative breast cancer44, as is hypomethylation of the CD133 (also known as PROM1) locus in glioblastoma stem cells⁴⁵. Although many studies suggest that DNMT3A promotes differentiation and acts as a tumour suppressor, DNMT3A and methylation status may have different consequences depending on the cellular context, as DNMT3A can be overexpressed in multiple

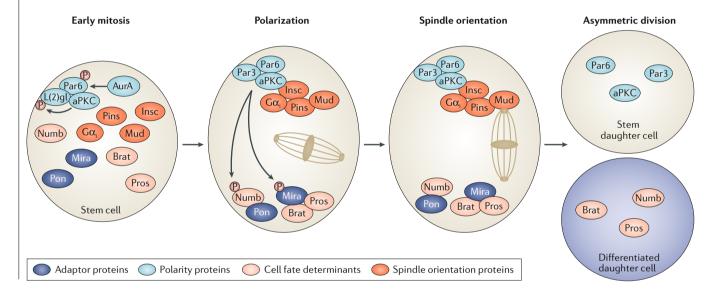
Box 1 | Asymmetric division

In *Drosophila melanogaster*, the clearest elucidation of the sequence of events leading to asymmetric division has come from studies of the neuroblast ^{191–193}. These cells undergo many rounds of asymmetric division in embryogenesis, generating one neuroblast and another ganglion mother cell that in turn gives rise to neurons and glia¹⁹⁴. Though these asymmetric divisions occur at distinct stages during development and adult life, similar mechanisms drive the balance of divisions, and homologues for most of the key regulators of these pathways exist in humans, suggesting conserved mechanisms of asymmetric division¹⁹³.

In the *Drosophila melanogaster* neuroblast, within a cell that will divide, atypical protein kinase C (aPKC) and Par6 are positioned at the apical cell cortex, a position inherited from a previous cell division. Here, they form a complex with Lethal(2) Giant Larvae Protein (L(2)gl), which prevents phosphorylation (P) of Numb by aPKC. Upon entry into mitosis, the kinase Aurora A (AurA) phosphorylates Par6, which in turn triggers aPKC to phosphorylate L(2)gl (see the figure). Phosphorylated L(2)gl is then released from the complex and replaced with Par3. Polarization results when aPKC phosphorylates Numb and the adaptor protein Miranda (Mira), restricting their localization to the basal region along with the adaptor

protein Partner of Numb (Pon). Miranda recruits Prospero (Pros) and Brain Tumour (Brat) to the basal membrane, allowing for the accumulation of these cell fate determinants by late prometaphase. The adaptor protein Inscuteable (Insc) then links the Par3–Par6–aPKC complex to the $G\alpha_i$ –Partner of Inscuteable (Pins) protein complex, which then interacts with Mushroom Body Defect (Mud), thereby linking the entire complex to the mitotic spindle and establishing its apical-basal orientation. Following cell division, the asymmetric inheritance of Numb acts to inhibit Notch signalling, and this in combination with the transcriptional activity of Prospromotes differentiation of the daughter cell.

The connection between aberrant asymmetric division and cancer was originally identified via a screen for genes that promote brain tumour development in *Drosophila melanogaster*⁶⁰⁻⁶³. Deletion of *l*(2)*gl, brat, prospero* and *numb* resulted in a loss of differentiation, uncontrolled cell proliferation and eventual development of brain tumours. Despite these early studies, progress in defining the link between division pattern and cancers in mammalian systems has been slow. However, over the past few years, emerging data have shown that this is an important regulator of cancer progression, and mutations in key regulators of this process are associated with oncogenesis^{35,60-65}.



Asymmetric division

A method of cellular diversification via differential segregation and inheritance of fate determinants leading to differently fated daughter cells. Controlled asymmetric division can be critically important during development but can become dysregulated during tumour initiation and progression.

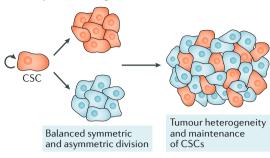
cancers, including breast, colon and liver cancers⁴⁶. Functionally, DNMT3A can also lead to a differentiation blockade such as that seen in hepatocellular carcinoma⁴⁷, and its deletion can block tumour progression in a model of colon cancer⁴⁸. Consistent with these findings, hypomethylating agents have been shown to promote differentiation and increase sensitivity to chemotherapies in some cancer cells⁴⁹. Given the context-specific impact of de novo DNA methylation, further work is clearly needed to define the programmes that differentially inhibit or promote tumorigenesis and to identify the cellular contexts most responsive to disruption of methyltransferase activity.

As a result of efforts to pharmacologically target epigenetic states, inhibitors of several broad-acting modulators such as enhancer of zeste homologue 2 (EZH2)⁵⁰, bromodomain-containing protein 4 (BRD4)⁵¹ and histone deacetylases (HDACs)⁵² have been shown to have a profound impact on tumour burden by promoting differentiation or by eroding stem cell programmes^{51,53–57}. Interestingly, perturbation of epigenetic programmes via

either gain or loss of histone acetylation using HDAC inhibitors or bromodomain inhibitors, respectively, can deplete CSCs^{55,58}. Similarly, loss or gain of DNA methylation via deletion or activation, respectively, of DNMT3A can trigger a collapse of oncogenic programmes and can impact stem cells preferentially relative to bulk tumour cells^{41,59}. The bidirectional nature of these dependencies suggests that cancer cells harbouring stem cell traits depend on tightly regulated networks, and either gain or loss of epigenetic modifications can be deleterious. Furthermore, because epigenetic regulators control large-scale programmes, targeting them may be particularly effective for perturbing the stem cell state in cancers.

Asymmetric division and stem cell fate. In addition to epigenetic programmes, a key way in which stem cell fate can be controlled is through asymmetric division, a post-translational mechanism critical for diversification through differential segregation and inheritance of proteins during cell division (BOX 1). Misappropriation of asymmetric division by oncogenic events can be a

Chronic phase or low grade



Acute phase or high grade

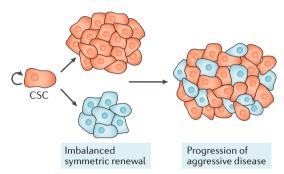


Fig. 3 | Asymmetric division in cancer. The disruption of asymmetric division is one way in which cancer may progress to an aggressive state. In low-grade cancers, symmetric renewal and asymmetric divisions are fairly balanced, resulting in both tumour heterogeneity and the maintenance of cancer stem cells (CSCs). However, in high-grade cancers, this balance may be shifted towards increased symmetric renewal, resulting in the expansion of the stem cell fraction, which may result in a more aggressive disease state. While imbalances in asymmetric division leading to the progression of cancer have been clearly demonstrated in haematological malignancies, there is evidence to suggest that disruption of asymmetric division can promote an aggressive state in some solid tumours as well.

potential force driving cancer. When asymmetric and symmetric divisions are balanced, tumours are heterogeneous, containing both stem cells and bulk cancer cells. However, when the balance is shifted towards symmetric division (renewal), this results in the expansion of CSCs that subsequently drive a more aggressive, undifferentiated state.

The connection between aberrant asymmetric division and cancer was originally identified in *Drosophila melanogaster*⁶⁰⁻⁶³, and has since been linked to mammalian cancers as well. The possibility that the differentiation arrest in aggressive cancers may be driven by disrupted asymmetric division was initially suggested by observations in haematological malignancies (FIG. 3; TABLE 1). While division patterns were not altered in chronic-phase CML, introduction of a second mutation leading to blast crisis CML triggered an imbalance favouring symmetric renewal⁶⁴. Mechanistically, this shift was driven by MSI³⁵, which repressed the pro-differentiation signal numb homologue (NUMB) to promote an aggressive undifferentiated state. Though dysregulation of asymmetric

division may result in a more aggressive cancer, the balance can be corrected: thus, both increased expression of NUMB or loss of MSI as well as inhibition of the dynein-binding protein lissencephaly 1 protein (LIS1; also known as PAFAH1B1), which leads to increased asymmetric division, served to halt the progression of aggressive myeloid disease in vivo^{35,65}.

As in leukaemia, a common theme in solid cancers involves disruption of NUMB leading to increased self-renewal. Receptor tyrosine-protein kinase ERBB2mutant breast cancer cells display increased symmetric renewal divisions66 triggered by symmetric NUMB inheritance. MSI signalling has also been implicated in other aggressive cancers such as pancreatic cancer, where it is an indicator of poor prognosis^{55,67}. p53 may also act in part by influencing symmetric renewal, with p53 loss reducing the frequency of asymmetric divisions and thus reducing differentiation in cells in the brain^{68,69}. These data suggest that hijacking asymmetric division can be a point of control for classic tumour suppressors and oncogenes and raise the possibility that enforced asymmetric division could be a strategy for controlling certain aggressive cancers.

Stem cell states in metastasis

Stem cell programmes and epithelial-mesenchymal **transition.** The conventional paradigm for metastasis was based originally on observations in breast cancer, and it postulated that cancer cells within primary tumours undergo epithelial-mesenchymal transition (EMT) and that this was necessary to enter circulation and transit to secondary sites⁷⁰. Although recent studies have raised doubts about the necessity of EMT during metastasis71,72, there is substantial evidence for a gradient of tumour cells^{73,74} expressing both epithelial and mesenchymal markers within the primary tumour, in circulation and at the secondary site75-77. However, in order for these mesenchymal cells to establish an epithelial tumour at the secondary site, genes responsible for maintaining a mesenchymal cell state must be switched off^{78,79}. These findings led to the idea that EMT occurs at the primary site and is followed by mesenchymal-epithelial transition at the secondary site for successful metastatic growth. This model bears striking parallels with the stem cell model, which postulates that a subpopulation of cells within the tumour has preferential capacity for driving tumour growth and regrowth at a new site and can effectively recreate tumour heterogeneity (FIG. 4).

The congruence between the stem cell and the EMT models of metastasis is supported by multiple observations showing that most disseminated tumour cells express stem cell markers^{55,80,81} and that functionally cells expressing stem cell markers like aldehyde dehydrogenase (ALDH) are highly enriched in their ability to form metastases^{82,83}. Consistent with the idea that the stem cell state is a critical part of EMT and metastatic potential, genome-wide analysis of cells undergoing EMT⁸⁴ and circulating tumour cells⁸⁵ revealed a remarkably congruent transcriptomic profile between these cells and primary CSCs⁸⁶. Circulating tumour cells isolated from patients with breast cancer⁸⁴ or from xenografts derived from patients with breast cancer⁸⁵ overexpress both EMT

Symmetric division

A method of cell division in which fate determinants are equivalently segregated. The resulting pair of daughter cells can either be undifferentiated (symmetric renewal) or differentiated daughter cells (symmetric commitment).

Tumour heterogeneity

Here, refers to the presence of functionally distinct malignant cells within a tumour. Heterogeneity can be driven by different genomic, transcriptomic or epigenetic landscapes.

Table 1 | Asymmetric division genes in cancer

Protein					
. rotem	Function in asymmetric division	Cancer type	Effect on asymmetric division	Dysregulation in cancer	Refs
LLGL1	Cell polarity	Leukaemia	Promotes asymmetric division	Decreased expression	195
NUMB	Cell fate	Leukaemia, colon cancer and breast cancer	Promotes differentiation	Decreased expression	35,66,199
MSI	Cell fate	Leukaemia	Promotes stemness	Increased expression	35
LIS1	Dynein binding and spindle orientation	Leukaemia	Promotes symmetric renewal	Critical for propagation of CSCs	65
TRIM3	Cell fate	Brain cancer	Promotes asymmetric division	Decreased expression	200
p53	Cell fate	Brain cancer, colon cancer and breast cancer	Promotes asymmetric division	Decreased expression	66,69,196
miR-34a	Cell fate	Colon cancer and brain cancer	Promotes differentiation (targets NOTCH)	Decreased expression	197–199
miR-146a	Cell fate	Colon cancer	Promotes symmetric renewal (targets NUMB)	Increased expression	199
lnc34a	Cell fate	Colon cancer	Promotes symmetric renewal (targets miR-34a)	Increased expression	201

CSCs, cancer stem cells; LLGL1, lethal(2) giant larvae protein homologue 1; LIS1, lissencephaly 1 protein; lnc, long non-coding RNA; miR, microRNA; MSI, RNA-binding protein Musashi homologue; NUMB, numb homologue; TRIM3, tripartite motif-containing protein 3.

markers (such as twist-related protein 1 (TWIST1), AKT2 and PI3K) and stem cell markers (such as ALDH, epithelial cell adhesion molecule (EPCAM), CD44, CD47 and MET) or exhibit stem cell properties such as chemoresistance. Genes associated with EMT are also highly expressed in CSCs⁸⁷; vimentin⁸⁸, transforming growth factor-β (TGFβ)89 and the transcription factors TWIST1 (REF. 90), zinc-finger protein SNAI1 and SNAI2 (REFS 91,92), and zinc-finger E-box-binding homeobox 1 (ZEB1) and ZEB2 (REF. 93) are enriched in and support the maintenance of stem cells from multiple cancers. Conventional EMT factors such as TWIST1, SNAI1, SNAI2 and ZEB1 can lead to acquisition of stem cell traits such as tumoursphere formation94,95 and activate expression of stem cell programmes driven by the transcription factors SOX2 and krüppel-like factor 4 (KLF4)%.

Metastatic stem cells. Although the discussion above strongly suggests that the stem cell state and EMT are in fact overlapping concepts that evolved in different fields, it is possible that these cells represent populations with substantial but not complete overlap. A 'metastatic stem cell'97 has been proposed as a population with increased metastatic capabilities that may not overlap with other stem cell properties such as therapy resistance or immediate capacity to propagate tumours. The metastatic stem cell could in fact be a subpopulation of stem cells or one that evolves with new mutations needed to trigger metastasis. For example, CD133+ pancreatic stem cells isolated from primary patient samples preferentially propagate tumours and are highly resistant to chemotherapy98. At the invasive front of tumour growth, CD133+ cells are enriched for CXC-chemokine receptor 4 (CXCR4) expression, and this double-positive population is more migratory than CD133+CXCR4- cells. Patients with more CD133+CXCR4+ cells had more metastatic disease, indicating the relevance of these cells for human disease98. A similar subpopulation of colorectal CSCs expressing CD26 was identified as the population responsible for liver metastasis and was predictive of distant metastasis in patients99.

Exposure to spatially distinct microenvironmental cues throughout the tumour could be one trigger for heterogeneity within CSCs. In this regard, CD133+CXCR4+ or CD133+CXCR4- cells may not be two distinct populations but might rather represent a gradient of stem cell programmes that are expressed at higher or lower levels in response to intracellular and intercellular signals. Emerging technologies using unbiased single-cell sequencing have independently supported the existence of intratumoural heterogeneity among cells with stem-like properties 100-103. New insights into the state of tumour cells driving metastasis and which programmes and cues may promote functionally distinct capacities will likely develop by applying these same unbiased technologies to metastatic tumour cells.

Stem cell states in therapy resistance

A major challenge in cancer therapy is the fact that not all cells within a tumour are equivalently sensitive to or effectively targeted by most therapies (FIG. 5a). In large part, the cells that are not eliminated contribute to residual disease and are the key drivers of cancer relapse. Thus, understanding the basis of differential sensitivity to drugs is critical to more efficient therapies and control of tumour growth. While some cytotoxic therapies have

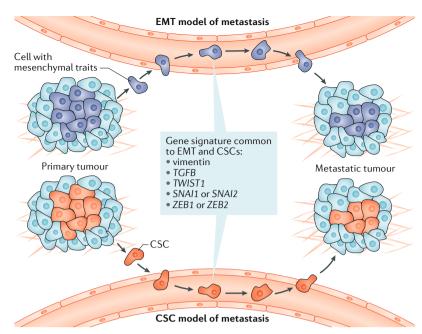


Fig. 4 | **Metastasis and cancer stem cells.** The classic epithelial–mesenchymal (EMT) model of metastasis (top) posits that the dissemination of cancer cells requires loss of epithelial cell traits commensurate with gain of mesenchymal cell traits (dark blue), which enables the cells to detach from the primary tumour and invade surrounding tissue, intravasate and survive in circulation and, finally, extravasate and localize to a distant metastatic site. Several genes (shown in the centre box) have been shown to drive EMT, and their expression serves as a marker of the process. Interestingly, cancer stem cells (CSCs) (bottom) are also enriched in disseminated tumour cells and express the EMT gene signature. Furthermore, the capacity for tumour propagation, which is required for establishment of a tumour at a distant site, is a salient feature of stem cells. The parallels between EMT cells and CSCs raise the possibility that they represent overlapping concepts.

been thought to directly induce mutations that can lead to acquired resistance^{104–106}, other studies have revealed pre-existing resistant clones within the tumour that drive tumour regrowth following therapy^{105–109}. Beyond genomic heterogeneity, it is becoming clear that epigenetic heterogeneity^{110,111} is also a key driver of differential sensitivity of cancer cells to multiple therapies. Such epigenetically driven resistance often depends on hijacked properties of normal stem cells such as the expression of drug transporters¹¹² (FIG. 5b), heightened DNA damage repair capacity¹¹³ (FIG. 5c) and recruitment of a protective niche (FIG. 5d,e)¹¹⁴.

Resistance to chemotherapy and radiotherapy. Cytotoxic drug efflux is frequently controlled by ATP-binding cassette (ABC) transporters, including the efflux pumps P glycoprotein 1 (also known as ABCB1) and ABC subfamily member 2 (ABCG2), which are highly expressed on normal and malignant haematopoietic and neural stem cells¹¹⁵⁻¹¹⁷. Because ABC transporters are generally promiscuous, they have the capacity to nonspecifically clear a range of toxic agents. Thus, cytotoxic chemotherapies are moderately successful at eliminating bulk tumour cells but leave behind aggressive stem cells that continue to express high levels of ABC transporters (FIG. 5b). In primary cell lines derived from patients with neuroblastoma, an ABCG2^{hi}ABCA3^{hi} side population of tumour cells is able to sustain long-term expansion ex vivo and rapidly clear

the cytotoxic drug mitoxantrone¹¹⁸. Interestingly, this population divides through asymmetric division to give rise to ABCG2^{hi}ABCA3^{hi} stem cells and more differentiated ABCG2^{low}ABCA3^{low} daughter cells, suggesting that drug pump expression is specifically inherited asymmetrically by the self-renewing daughter cell.

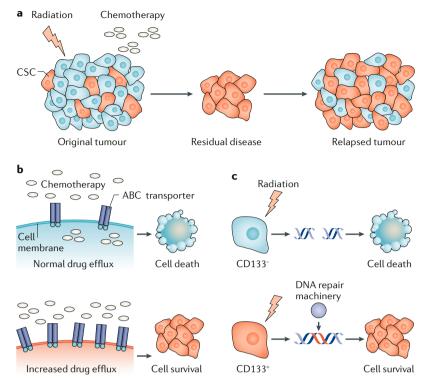
Resistance to radiation has been well studied, and its links to stem cell traits are perhaps best explored in glioblastoma, in which radiation is a standard of care. While radiotherapy improves overall survival and quality of life, most patients relapse even following full remission¹¹⁹. CD133⁺ cancer cells, a key population driving tumour growth in human disease¹²⁰, are highly enriched following radiation in vitro and in patient xenografts¹²¹. This enrichment appears to be driven by the preferential ability of the stem cell population to repair DNA damage (FIG. 5c) by activating checkpoint kinase 1 (CHK1) and CHK2. While preclinical studies indicated that these stem cells could be radio-sensitized with CHK1 and CHK2 inhibitors¹²¹, this therapeutic approach failed in trials owing to high toxicity¹²². Recent studies suggest that glioma stem cells also rely on PCNA-associated factor (PAF)-driven translesion DNA synthesis for preferential survival following radiation¹²³: pharmacologic inhibition of translesion DNA synthesis leads to radiosensitization and depletion of glioma stem cells and thus represents a novel therapeutic approach for patients with glioblastoma. Efforts to identify new strategies to erode programmes that enable enhanced DNA repair in stem cells remain critical to improving the durability of non-targeted as well as some targeted therapies.

Targeted and immunotherapies. In the past few decades, the greatest strides in molecularly targeted therapies have been led by the discovery of imatinib, the first tyrosine kinase inhibitor. Imatinib effectively blocks BCR-ABL activity in CML and leads to remarkably effective prevention of CML progression¹²⁴. However, among patients with CML and minimal evidence of disease, approximately half relapsed within the first year of imatinib withdrawal¹²⁵. This relapse was found to be driven by residual disease comprising leukaemia stem cells126-128. Although imatinib is effective in blocking BCR-ABL in the stem cell fraction¹²⁹, CML stem cells are insensitive to imatinib because they are not addicted to BCR-ABL. Instead, resistant leukaemia stem cells activate several alternative signals to enable survival and renewal, including β-catenin, SMO and arachidonate 5-lipoxygenase (ALOX5)130-133. These broad patterns have also been observed in lung cancer, in which therapies targeting EGFR mutations lead to enrichment of stem-like cells that are dependent on NOTCH3 (REFS^{134,135}), and this resistance can be overcome by inhibiting Notch signalling¹³⁶. This provided an early and important example of drug resistance without the evolution of any new mutations and is one of the best examples of a disease in which the stem cell fraction is the key contributor to residual disease.

With the advent of new cancer therapies exploiting the innate ability of the immune system to track and kill cancer cells^{137–140}, understanding resistance to such therapies has become an increasing focus, and stem cell signals appear to be relevant in this context. A machine-learning

Side population

A small population of cells detected via flow cytometry that has increased dye efflux, a property that is associated with an increased expression of drug transporters. Functionally, the side population is enriched for cells with the ability to self-renew and differentiate. As these are key features of stem cells, the side population has traditionally been found to be enriched in stem cells and cancer stem cells.



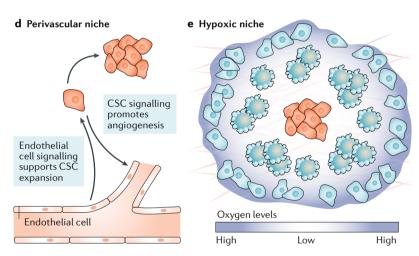


Fig. 5 | Therapy resistance in cancer stem cells. a | Cytotoxic agents such as radiation and chemotherapy are commonly used to treat cancer, efficiently targeting bulk cancer cells (blue cells) but not cancer stem cells (CSCs) (orange cells). The residual disease can be enriched in stem cell populations that can drive a more aggressive disease, triggering recurrence. b | Stem cell properties are commonly hijacked in cancer. One such property is increased drug efflux. Chemotherapeutic agents target bulk cancer cells with normal levels of drug efflux, resulting in cell death (top). In CSCs, higher expression of ATP-binding cassette (ABC) transporters can increase drug efflux capacity, increasing cell survival (bottom). c | Enhanced DNA repair can also be hijacked in cancer. In glioblastoma, radiation generates unrepaired double strand breaks in CD133-bulk cancer cells, leading to cell death (top). In CD133+ CSCs (bottom), the DNA damage checkpoint is activated, allowing for repair that leads to increased cell survival. **d** CSCs utilize the tumour microenvironment for increased survival. In brain tumours, the endothelial cells of the perivascular niche promote the survival of CSCs. Endothelial cell signalling supports the stem cell properties of the cancer, which allows CSC expansion. CSCs can promote angiogenesis by secreting factors such as vascular endothelial growth factor (VEGF) and stromal cell-derived factor 1 (SDF1). e | Hypoxic environments can support CSCs. Although hypoxia (represented by the descending oxygen gradient shown in blue) induces some cell death within the tumour, it also promotes stem cell expansion and triggers expression of genes that promote therapy resistance.

algorithm used to identify epigenetic and transcriptomic signatures revealed that a stem-high, undifferentiated tumour landscape is associated with lower immune infiltration and downregulated programmed cell death 1 ligand 1 (PD-L1) signalling¹⁴¹, are characteristics that predict a poor response to immunotherapy^{142,143}. This link is supported by earlier data in melanoma, in which tumours with high T cell infiltration responded to immune checkpoint inhibitors¹⁴³, and T cell infiltration was found only in tumours with low WNT-β-catenin signalling¹⁴⁴. These data suggest that CSC signals can alter the tumour microenvironment by directly modulating tumour infiltrating lymphocytes. Bladder CSCs also modulated tumour infiltrating lymphocytes by producing inflammatory mediators like interleukin-6 (IL-6) and IL-8, which led to infiltration of pro-tumorigenic myeloid cells¹⁴⁵. In many ways, these studies exemplify the interplay between stem cells and the stem cell niche and highlight the importance of mapping the complex interactions CSCs make in vivo that influence the rise of resistance.

The microenvironment in resistance. While intrinsic mechanisms of therapy resistance have been more frequently linked to increased survival of stem cells, emerging studies suggest that the microenvironment may be equally critical. In brain tumours, endothelial cells have been shown to interact closely with stem-like cells and secrete factors that support maintenance of stem cell traits^{114,146–150} (FIG. 5d). For example, endothelial cells can induce expression of stem cell programmes in glioma cells by secreting nitric oxide to promote Notch signalling¹⁴⁹ or by secreting the CD44 ligand osteopontin¹⁵⁰. By contrast, endothelial cell inhibition through the use of the vascular endothelial growth factor (VEGF) inhibitor bevacizumab¹⁵¹ may also promote stem-like characteristics in non-stem cells through anti-VEGF-triggered hypoxia, which can block CSC differentiation^{152,153} (FIG. 5e). As an example, hypoxia triggers β-interferon gene positive regulatory domain I-binding factor (BLIMP1; also known as PRDM1) expression in pancreatic cancer cells¹⁵⁴, which subsequently activates EMT genes associated with therapy resistance. These examples highlight the challenges of interpreting studies involving signals from the tumour microenvironment, as they can be pleiotropic and involve multiple cell types.

In addition to endothelial cells, recent studies have highlighted important roles for other niche components in therapy resistance. Non-stem cells help maintain a pool of CSCs by secreting supportive signals such as WNT in lung adenocarcinoma¹⁵⁵ and brain-derived neurotrophic factor (BDNF) in glioblastoma¹⁵⁶. Analysis of cancer-associated fibroblasts from breast cancer samples before and after chemotherapy revealed an enrichment of fibroblasts in therapy-resistant tumours¹⁵⁷. This population was not only resistant to chemotherapy but also created a therapy-resistant niche by closely interacting with CSCs and secreting factors such as IL-6 and IL-8 that promoted stem cell survival¹⁵⁷. Fibroblasts have also been shown to promote CSC survival and expansion in non-small-cell lung cancer¹⁵⁸, basal cell carcinoma¹⁵⁹ and colorectal cancer¹⁶⁰.

Although the microenvironment is generally thought to be particularly important for therapy resistance in solid

Nutlin-3a

A drug that inhibits the interaction of E3 ubiquitin-protein ligase MDM2 and p53 and is most effective on cells with wild-type p53 expression. Once released, wild-type p53 can induce cell cycle arrest and apoptosis.

cancers, emerging evidence shows that leukaemia cells, which are generally considered to be highly motile, may in fact share this dependency. For example, genetic loss of CD98, a hub for integrin signalling, triggers defects in interactions of acute myeloid leukaemia (AML) stem cells with endothelial cells and leads to their depletion¹⁶¹. Similarly, tetraspanin 3 (TSPAN3) loss blocked AML localization to CXC-chemokine ligand 12 (CXCL12)-rich bone marrow regions and led to impaired leukaemia and AML stem cell growth¹⁶². In addition to myeloid leukaemia, T-ALL-initiating cells are dependent on CXCR4-mediated cell motility for survival, and microenvironment-derived CXCL12 is essential for CXCR4 activation¹⁶³. In human B cell precursor-ALL and T-ALL, long-term dormant cells are preferentially therapy resistant when associated with microenvironmental cells, suggesting that the microenvironment can drive therapy resistance¹⁶⁴. These studies highlight the importance of niche signals for leukaemia stem cell homing, proliferation and survival.

New technologies

The recent development of culture conditions that support long-term expansion of normal and neoplastic organoids165,166 has provided a new platform for identifying drivers of therapy resistance and improving prediction of good responders. Importantly, patient-derived organoid cultures from colorectal cancer¹⁶⁷, pancreatic cancer^{168,169}, breast cancer¹⁷⁰, liver cancer¹⁷¹ and bladder cancer¹⁷² have been shown to retain genetic mutations present in the parental tumour sample. As expected, colorectal cancer organoids with wild-type p53 responded well to nutlin-3a, and those with activating mutations in the WNT pathway were sensitive to WNT inhibitors 167. Additionally, in vitro drug screens using patient-derived organoids recapitulated in vivo xenograft drug response^{170,172}, which supports the robust nature of this system for accurately predicting therapy response. Interestingly, much of the variability in therapy response in tumour organoids could only be marginally explained by mutation burden^{167,172}, suggesting diverse mechanisms of therapy resistance that reflect patient diversity. This was supported by longitudinal tracking of chemotherapy response in pancreatic cancer patients and their matched tumour organoids169: organoids that were markedly responsive or resistant to specific chemotherapies coincided with patient outcome. Moreover, parallel transcriptomic analysis led to the identification of transcriptional signatures that correlated with patient response¹⁶⁹. Because tumour organoids are specifically derived from CSCs in the colon¹⁶⁷ and the pancreas (N.K.L., T.R. and N. Rajbhandari, unpublished observations), the studies discussed above provide a unique platform for measuring the drug responsiveness of a heterogenous population that is sustained by stem cell programmes. Thus, drug-sensitive organoid signatures provide a unique perspective on intertumoural stem cell heterogeneity and may allow us to better predict vulnerabilities.

Perspectives

The discussion above provides a view into how stem cell programmes can enable cancer initiation, therapy resistance and metastasis. The compelling biology in this rapidly moving field has already led to the development

of agents targeting stem cell signals that have emerged as an important new class of differentiation therapies. Among these, the SMO antagonists, which inhibit the Hedgehog pathway, are furthest along, and are approved for use in the treatment of advanced basal cell carcinoma. These antagonists have been in trials for several other cancers as well, including medulloblastoma and lung cancer¹⁷³⁻¹⁸⁰ on the basis of findings from studies identifying the importance of the Hedgehog pathway in these cancers^{133,181–183}. The Notch pathway has been inhibited using y-secretase inhibitors, which prevent cleavage of NOTCH, though are not specific to Notch signalling³². More recently, anti-DLL4 monoclonal antibodies, which more specifically target the Notch pathway, have also been developed and are in trials for multiple advanced malignancies, including metastatic colorectal cancer and ovarian cancer¹⁸⁴. The development of WNT inhibitors, while critical given its extensive mutation in colon cancer and activation in multiple other cancers, has been a more challenging undertaking¹⁸⁵. However, the development of a CBP-β-catenin antagonist (PRI-724), which interferes with the binding of β -catenin with CBP and not p300 (REF. 186), has allowed clinical testing of WNT pathway inhibition in advanced myeloid malignancies. Additional trials have tested the impact of inhibiting the WNT pathway at the level of WNT secretion or receptor binding using an anti-frizzled 7 (FZD7) receptor monoclonal antibody (vantictumab)187, a WNT ligand antagonist (ipafricept)¹⁸⁸ or a protein-serine O-palmitoleoyltransferase porcupine inhibitor (LGK974)¹⁸⁹ in pancreatic and breast cancers.

At a broader level, it is worth considering the fact that despite the intense focus on identifying key signalling events and targeting these as potential strategies for therapeutic intervention, the rate of failure in trials remains high. It is likely that many drugs could be very effective, but inefficient delivery and trials in advanced stage disease likely reduce their impact on tumour growth. Improving methods of delivery through nanoparticle or lipid-mediated delivery, antibody-drug conjugate strategies and local delivery efforts represents a crucial area to explore to improve outcomes. The issue of early intervention has important ramifications for treatment outcomes in general. Among targeted therapies, imatinib is extraordinary in leading to remarkable long-term remissions that have allowed a majority of patients to live normal lives. Though usually considered a poster child of targeted therapies, the success of imatinib may have more to do with it being a true early intervention, as CML can be detected in the indolent and benign chronic phase, and imatinib is far less successful in controlling the disease as CML progresses into blast crisis 190. This highlights the need for a greater focus on early detection methods and raises the possibility that strategies to detect stem cell signatures could be useful as indicators of disease progression. Combining the development of innovative early detection tools with an understanding of the signals that drive benign disease to a more malignant phase would enable effective early intervention and provide a more balanced approach to controlling cancer.

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- Hunger, S. P., Winick, N. J., Sather, H. N. & Carroll, W. L. Therapy of low-risk subsets of childhood acute lymphoblastic leukemia: when do we say enough? *Pediatr. Blood Cancer* 45, 876–880 (2005).
- Hodgson, D. C., Hudson, M. M. & Constine, L. S. Pediatric hodgkin lymphoma: maximizing efficacy and minimizing toxicity. Semin. Radiat. Oncol. 17, 230–242 (2007).
- Barker, N. et al. Crypt stem cells as the cells-of-origin of intestinal cancer. *Nature* 457, 608–611 (2009).
- Gibson, P. et al. Subtypes of medulloblastoma have distinct developmental origins. *Nature* 468, 1095–1099 (2010).
- Lapouge, G. et al. Identifying the cellular origin of squamous skin tumors. *Proc. Natl Acad. Sci. USA* 108, 7431–7436 (2011).
- Xu, X. et al. Evidence for type II cells as cells of origin of K-Ras-induced distal lung adenocarcinoma. *Proc. Natl Acad. Sci. USA* 109, 4910–4915 (2012).
- Reya, T., Morrison, S. J., Clarke, M. F. & Weissman, I. L. Stem cells, cancer, and cancer stem cells. *Nature* 414, 105–111 (2001).
- Daley, G. Q., Van Etten, R. A. & Baltimore, D. Induction of chronic myelogenous leukemia in mice by the P210bcr/abl gene of the Philadelphia chromosome. Science 247, 824–830 (1990).
- Science **247**, 824–830 (1990).

 9. Whang, J., Frei, E. 3rd, Tjio, J. H., Carbone, P. P. & Brecher, C. The distribution of the Philadelphia chromosome in patients with chronic myelogenous leukemia. *Blood* **22**, 664–673 (1963).
- Krivtsov, A. V. et al. Cell of origin determines clinically relevant subtypes of MLL-rearranged AML. *Leukemia* 27, 852–860 (2013).
- Shlush, L. I. et al. Identification of pre-leukaemic haematopoietic stem cells in acute leukaemia. *Nature* 506, 328–333 (2014).
- Blaas, L. et al. Lgr6 labels a rare population of mammary gland progenitor cells that are able to originate luminal mammary tumours. *Nat. Cell Biol.* 18, 1346–1356 (2016).
- Zhao, C. et al. Loss of beta-catenin impairs the renewal of normal and CML stem cells in vivo. *Cancer Cell* 12, 528–541 (2007).
- Latil, M. et al. Cell-type-specific chromatin states differentially prime squamous cell carcinoma tumorinitiating cells for epithelial to mesenchymal transition. Cell Stem Cell 20, 191–204 (2017).
- Cell Stem Cell 20, 191–204 (2017).
 Alcantara Llaguno, S. R. et al. Adult lineage-restricted CNS progenitors specify distinct glioblastoma subtypes. Cancer Cell 28, 429–440 (2015).
- Alcantara Llaguno, S. et al. Malignant astrocytomas originate from neural stem/progenitor cells in a somatic tumor suppressor mouse model. *Cancer Cell* 15, 45–56 (2009).
- Yang, Z. J. et al. Medulloblastoma can be initiated by deletion of Patched in lineage-restricted progenitors or stem cells. *Cancer Cell* 14, 135–145 (2008).
- Schuller, U. et al. Acquisition of granule neuron precursor identity is a critical determinant of progenitor cell competence to form Shh-induced medulloblastoma. *Cancer Cell* 14, 123–134 (2008).
- Xie, W. et al. DNA methylation patterns separate senescence from transformation potential and indicate cancer risk. *Cancer Cell* 33, 309–321 (2018).
- Yu, Y. et al. Targeting the senescence-overriding cooperative activity of structurally unrelated H3K9 demethylases in melanoma. *Cancer Cell* 33, 322–336 (2018).
- Kaufman, C. K. et al. A zebrafish melanoma model reveals emergence of neural crest identity during melanoma initiation. Science 351, aad2197 (2016). This study illustrates that the epigenetic state can act as a determining factor for the cell of origin in a zebrafish model of melanoma.
- Gilbert, N. et al. Chromatin architecture of the human genome: gene-rich domains are enriched in open chromatin fibers. *Cell* 118, 555–566 (2004).
- Hoadley, K. A. et al. Cell-of-origin patterns dominate the molecular classification of 10,000 tumors from 33 types of cancer. *Cell* 173, 291–304 (2018).
- Vaz, M. et al. Chronic cigarette smoke-induced epigenomic changes precede sensitization of bronchial epithelial cells to single-step transformation by KRAS mutations. Cancer Cell 32, 360–376 (2017).
- Kinzler, K. W. & Vogelstein, B. Lessons from hereditary colorectal cancer. *Cell* 87, 159–170 (1996).
- Cho, Y. J. et al. Integrative genomic analysis of medulloblastoma identifies a molecular subgroup that drives poor clinical outcome. *J. Clin. Oncol.* 29, 1424–1430 (2011).

- Hahn, H. et al. Mutations of the human homolog of Drosophila Patched in the nevoid basal cell carcinoma syndrome. *Cell* 85, 841–851 (1996).
- Ellisen, L. W. et al. TAN-1, the human homolog of the Drosophila Notch gene, is broken by chromosomal translocations in T lymphoblastic neoplasms. *Cell* 66, 649–661 (1991)
- Gonzalez, M. E. et al. EZH2 expands breast stem cells through activation of NOTCH1 signaling. *Proc. Natl Acad. Sci. USA* 111, 3098–3103 (2014).
- Miyamoto, Y. et al. Notch mediates TGF alpha-induced changes in epithelial differentiation during pancreatic tumorigenesis. *Cancer Cell* 3, 565–576 (2003).
- Valencia, A. et al. Wnt signaling pathway is epigenetically regulated by methylation of Wnt antagonists in acute myeloid leukemia. *Leukemia* 23, 1658–1666 (2009).
- Yuan, X. et al. Notch signaling: an emerging therapeutic target for cancer treatment. *Cancer Lett.* 369, 20–27 (2015).
- Ashihara, E., Takada, T. & Maekawa, T. Targeting the canonical Wnt/beta-catenin pathway in hematological malignancies. *Cancer Sci.* 106, 665–671 (2015).
- Di Giacomo, D. et al. Blast crisis Ph+ chronic myeloid leukemia with NUP98/HOXA13 up-regulating MSI2. Mol. Cytogenet. 7, 42 (2014).
- Ito, T. et al. Regulation of myeloid leukaemia by the cell-fate determinant Musashi. *Nature* 466, 765–768 (2010).
- Kharas, M. G. et al. Musashi-2 regulates normal hematopoiesis and promotes aggressive myeloid leukemia. *Nat. Med.* 16, 903–908 (2010).
- Yamashita, Y. et al. Array-based genomic resequencing of human leukemia. *Oncogene* 29, 3723–3731 (2010).
- Ley, T. J. et al. DNMT3A mutations in acute myeloid leukemia. N. Engl. J. Med. 363, 2424–2433 (2010).
- Challen, G. A. et al. Dnmt3a is essential for hematopoietic stem cell differentiation. *Nat. Genet.* 44, 23–31 (2011).
- Tadokoro, Y., Ema, H., Okano, M., Li, E. & Nakauchi, H. De novo DNA methyltransferase is essential for selfrenewal, but not for differentiation, in hematopoietic stem cells. J. Exp. Med. 204, 715–722 (2007).
- Mayle, A. et al. Dnmt3a loss predisposes murine hematopoietic stem cells to malignant transformation. *Blood* 125, 629–638 (2015).
- Hajkova, H. et al. Decreased DNA methylation in acute myeloid leukemia patients with DNMT3A mutations and prognostic implications of DNA methylation. *Leuk. Res.* 36, 1128–1133 (2012).
- Yan, X. J. et al. Exome sequencing identifies somatic mutations of DNA methyltransferase gene DNMT3A in acute monocytic leukemia. *Nat. Genet.* 43, 309–315 (2011).
- Kagara, N. et al. Epigenetic regulation of cancer stem cell genes in triple-negative breast cancer. *Am. J. Pathol.* 181, 257–267 (2012).
 Gopisetty, G., Xu, J., Sampath, D., Colman, H. &
- Gopisetty, G., Xu, J., Sampath, D., Colman, H. & Puduvalli, V. K. Epigenetic regulation of CD133/ PROM1 expression in glioma stem cells by Sp1/myc and promoter methylation. *Oncogene* 32, 3119–3129 (2013)
- 3119–3129 (2013).
 46. Zhang, W. & Xu, J. DNA methyltransferases and their roles in tumorigenesis. *Biomark. Res.* 5, 1 (2017).
- Martin, M. et al. Dynamic imbalance between cancer cell subpopulations induced by transforming growth factor beta (ΤGF-β) is associated with a DNA methylome switch. BMC Genomics 15, 435 (2014).
- Weis, B. et al. Inhibition of intestinal tumor formation by deletion of the DNA methyltransferase 3a. Oncogene 34, 1822–1830 (2015).
- Tsai, H. C. et al. Transient low doses of DNAdemethylating agents exert durable antitumor effects on hematological and epithelial tumor cells. *Cancer Cell* 21, 430–446 (2012).
- Kondo, Y. Targeting histone methyltransferase EZH2 as cancer treatment. J. Biochem. 156, 249–257 (2014)
- Žuber, J. et al. RNAi screen identifies Brd4 as a therapeutic target in acute myeloid leukaemia. Nature 478, 524–528 (2011).
 This study identifies the epigenetic regulator BRD4 as critical in the maintenance of CSCs in AML.
- Riggs, M. G., Whittaker, R. G., Neumann, J. R. & Ingram, V. M. n-Butyrate causes histone modification in HeLa and Friend erythroleukaemia cells. *Nature* 268, 462–464 (1977).
- Fiskus, W. et al. Highly active combination of BRD4 antagonist and histone deacetylase inhibitor against human acute myelogenous leukemia cells. Mol. Cancer Ther. 13, 1142–1154 (2014).

- Pei, Y. et al. HDAC and PI3K antagonists cooperate to inhibit growth of MYC-driven medulloblastoma. Cancer Cell 29, 311–323 (2016).
- 55. Fox, R. G. et al. Image-based detection and targeting of therapy resistance in pancreatic adenocarcinoma. Nature 534, 407–411 (2016). This study shows that the stem cell fate determinant MSI is a critical mediator of
- pancreatic cancer progression and lethality.
 66. Mazur, P. K. et al. Combined inhibition of BET family proteins and histone deacetylases as a potential epigenetics-based therapy for pancreatic ductal adenocarcinoma. *Nat. Med.* 21, 1163–1171 (2015).
- Shu, S. et al. Response and resistance to BET bromodomain inhibitors in triple-negative breast cancer. *Nature* **529**, 413–417 (2016).
- Zhang, B. et al. Effective targeting of quiescent chronic myelogenous leukemia stem cells by histone deacetylase inhibitors in combination with imatinib mesylate. *Cancer Cell* 17, 427–442 (2010).
- Pathania, R. et al. DNMT1 is essential for mammary and cancer stem cell maintenance and tumorigenesis. *Nat. Commun.* 6, 6910 (2015).
- Bello, B., Reichert, H. & Hirth, F. The brain tumor gene negatively regulates neural progenitor cell proliferation in the larval central brain of Drosophila. *Development* 133, 2639–2648 (2006).
 Gateff, E. Malignant neoplasms of genetic origin in
- Gateff, E. Malignant neoplasms of genetic origin in Drosophila melanogaster. *Science* 200, 1448–1459 (1978)
- Betschinger, J., Mechtler, K. & Knoblich, J. A. Asymmetric segregation of the tumor suppressor brat regulates self-renewal in Drosophila neural stem cells. *Cell* 124, 1241–1253 (2006).
- Bowman, S. K., Neumuller, R. A., Novatchkova, M., Du, Q. & Knoblich, J. A. The Drosophila NuMA Homolog Mud regulates spindle orientation in asymmetric cell division. *Dev. Cell* 10, 731–742 (2006).
- 64. Wu, M. et al. Imaging hematopoietic precursor division in real time. *Cell Stem Cell* 1, 541–554 (2007).
- Zimdahl, B. et al. Lis1 regulates asymmetric division in hematopoietic stem cells and in leukemia. *Nat. Genet.* 46, 245–252 (2014).
- Cicalese, A. et al. The tumor suppressor p53 regulates polarity of self-renewing divisions in mammary stem cells. *Cell* 138, 1083–1095 (2009).
- Sheng, W. et al. Musashi2 promotes the development and progression of pancreatic cancer by down-regulating Numb protein. *Oncotarget* 8, 14359–14373 (2016).
- Shen, Q., Zhong, W., Jan, Y. N. & Temple, S. Asymmetric Numb distribution is critical for asymmetric cell division of mouse cerebral cortical stem cells and neuroblasts. *Development* 129, 4843–4853 (2002).
- Sugiarto, S. et al. Asymmetry-defective oligodendrocyte progenitors are glioma precursors. *Cancer Cell* 20, 328–340 (2011).
- Thiery, J. P. Epithelial-mesenchymal transitions in tumour progression. *Nat. Rev. Cancer* 2, 442–454 (2002).
- Zheng, X. et al. Epithelial-to-mesenchymal transition is dispensable for metastasis but induces chemoresistance in pancreatic cancer. *Nature* 527, 525–530 (2015).
- Fischer, K. R. et al. Epithelial-to-mesenchymal transition is not required for lung metastasis but contributes to chemoresistance. *Nature* 527, 472–476 (2015).
- Jolly, M. K., Waré, K. E., Gilja, S., Somarelli, J. A. & Levine, H. EMT and MET: necessary or permissive for metastasis? *Mol. Oncol.* 11, 755–769 (2017).
- Aiello, N. M. et al. EMT subtype influences epithelial plasticity and mode of cell migration. *Dev. Cell* 45, 681–695 (2018).
- Sampson, V. B. et al. Wilms' tumor protein induces an epithelial-mesenchymal hybrid differentiation state in clear cell renal cell carcinoma. *PLOS ONE* 9, e102041 (2014).
- Schliekelman, M. J. et al. Molecular portraits of epithelial, mesenchymal, and hybrid states in lung adenocarcinoma and their relevance to survival. *Cancer Res.* 75, 1789–1800 (2015).
 Grosse-Wilde, A. et al. Stemness of the hybrid
- Grosse-Wilde, A. et al. Stemness of the hybrid epithelial/mesenchymal state in breast cancer and its association with poor survival. *PLOS ONE* 10, e0126522 (2015).
- Ocana, O. H. et al. Metastatic colonization requires the repression of the epithelial-mesenchymal transition inducer Prrx1. Cancer Cell 22, 709–724 (2012).
- 79. Tsai, J. H., Donaher, J. L., Murphy, D. A., Chau, S. & Yang, J. Spatiotemporal regulation of

- epithelial-mesenchymal transition is essential for squamous cell carcinoma metastasis. *Cancer Cell* **22**, 725–736 (2012).
- Balic, M. et al. Most early disseminated cancer cells detected in bone marrow of breast cancer patients have a putative breast cancer stem cell phenotype. *Clin. Cancer Res.* 12, 5615–5621 (2006).
- Grillet, F. et al. Circulating tumour cells from patients with colorectal cancer have cancer stem cell hallmarks in ex vivo culture. *Gut* 66, 1802–1810 (2017).
- Charafe-Jauffret, E. et al. Breast cancer cell lines contain functional cancer stem cells with metastatic capacity and a distinct molecular signature. Cancer Res. 69, 1302–1313 (2009).
- Dieter, S. M. et al. Distinct types of tumor-initiating cells form human colon cancer tumors and metastases. *Cell Stem Cell* 9, 357–365 (2011).
- Aktas, B. et al. Stem cell and epithelial-mesenchymal transition markers are frequently overexpressed in circulating tumor cells of metastatic breast cancer patients. *Breast Cancer Res.* 11, R46 (2009).
- Baccelli, I. et al. Identification of a population of blood circulating tumor cells from breast cancer patients that initiates metastasis in a xenograft assay. Nat. Biotechnol. 31, 539–544 (2013).
- Al-Hajj, M., Wicha, M. S., Benitò-Hernandez, A., Morrison, S. J. & Clarke, M. F. Prospective identification of tumorigenic breast cancer cells. *Proc. Natl Acad. Sci. USA* 100, 3983–3988 (2003).
- Polyak, K. & Weinberg, R. A. Transitions between epithelial and mesenchymal states: acquisition of malignant and stem cell traits. *Nat. Rev. Cancer* 9, 265–273 (2009).
- Raimondi, C. et al. Epithelial-mesenchymal transition and stemness features in circulating tumor cells from breast cancer patients. *Breast Cancer Res. Treat.* 130, 449–455 (2011).
- Shipitsin, M. et al. Molecular definition of breast tumor heterogeneity. *Cancer Cell* 11, 259–273 (2007)
- Beck, B. et al. Different levels of Twist1 regulate skin tumor initiation, stemness, and progression. *Cell Stem Cell* 16, 67–79 (2015).
- Fan, F. et al. Overexpression of snail induces epithelialmesenchymal transition and a cancer stem cell-like phenotype in human colorectal cancer cells. Cancer Med. 1, 5–16 (2012).
- Kurrey, N. K. et al. Snail and slug mediate radioresistance and chemoresistance by antagonizing p53-mediated apoptosis and acquiring a stem-like phenotype in ovarian cancer cells. Stem Cells 27, 2059–2068 (2009).
- Burk, U. et al. A reciprocal repression between ZEB1 and members of the miR-200 family promotes EMT and invasion in cancer cells. EMBO Rep. 9, 582–589 (2008).
- 94. Mani, S. A. et al. The epithelial-mesenchymal transition generates cells with properties of stem cells. *Cell* **133**, 704–715 (2008). This study links EMT and stem cells by reporting that EMT leads to the acquisition of stem cell
 - this study links EMT and stem cells by reporting that EMT leads to the acquisition of stem cell traits, while stem cells themselves express markers of EMT.
- Morel, A. P. et al. Generation of breast cancer stem cells through epithelial-mesenchymal transition. *PLOS ONE* 3, e2888 (2008).
- Wellner, U. et al. The EMT-activator ZEB1 promotes tumorigenicity by repressing stemness-inhibiting microRNAs. Nat. Cell Biol. 11, 1487–1495 (2009).
- Oskarsson, T., Batlle, E. & Massague, J. Metastatic stem cells: sources, niches, and vital pathways. Cell Stem Cell 14, 306–321 (2014).
- Hermann, P. C. et al. Distinct populations of cancer stem cells determine tumor growth and metastatic activity in human pancreatic cancer. *Cell Stem Cell* 1, 313–323 (2007).
- Pang, R. et al. A subpopulation of CD26+ cancer stem cells with metastatic capacity in human colorectal cancer. *Cell Stem Cell* 6, 603–615 (2010).
 Patel, A. P. et al. Single-cell RNA-seq highlights
- Patel, A. P. et al. Single-cell RNA-seq highlights intratumoral heterogeneity in primary glioblastoma. Science 344, 1396–1401 (2014).
- Tirosh, I. et al. Single-cell RNA-seq supports a developmental hierarchy in human oligodendroglioma. Nature 539, 309–313 (2016).
- 102. Bakker, B. et al. Single-cell sequencing reveals karyotype heterogeneity in murine and human malignancies. *Genome Biol.* 17, 115 (2016).
- Turner, K. M. et al. Extrachromosomal oncogene amplification drives tumour evolution and genetic heterogeneity. *Nature* 543, 122–125 (2017).
- 104. Pao, W. et al. Acquired resistance of lung adenocarcinomas to gefitinib or erlotinib is associated

- with a second mutation in the EGFR kinase domain. *PLOS Med.* **2**, e73 (2005).
- PLOS Med. 2, e73 (2005).
 105. Ding, L. et al. Clonal evolution in relapsed acute myeloid leukaemia revealed by whole-genome sequencing. Nature 481, 506–510 (2012).
- Patch, A. M. et al. Whole-genome characterization of chemoresistant ovarian cancer. *Nature* 521, 489–494 (2015).
- Lee, M. C. et al. Single-cell analyses of transcriptional heterogeneity during drug tolerance transition in cancer cells by RNA sequencing. Proc. Natl Acad. Sci. USA 111, E4726–E4735 (2014).
- Kurtova, A. V. et al. Blocking PGE2-induced tumour repopulation abrogates bladder cancer chemoresistance. *Nature* 517, 209–213 (2015).
- 109. Kim, C. et al. Chemoresistance evolution in triple-negative breast cancer delineated by single-cell sequencing. Cell 173, 879–893 (2018).
 Using single-cell sequencing, this study shows that therapy resistance in triple-negative breast cancer is driven by a population of pre-existing clones and not through the progressive accumulation of oncogenic mutations.
- Sharma, S. V. et al. A chromatin-mediated reversible drug-tolerant state in cancer cell subpopulations. *Cell* 141, 69–80 (2010).
- Hata, A. N. et al. Tumor cells can follow distinct evolutionary paths to become resistant to epidermal growth factor receptor inhibition. *Nat. Med.* 22, 262–269 (2016).
 - This study demonstrates that the epigenetic state can confer a drug-tolerant state, which then allows for genomic evolution leading to drug resistance.
- Dean, M. Cancer stem cells: implications for cancer causation and therapy resistance. *Discov. Med.* 5, 278–282 (2005).
- 113. Blanpain, C., Mohrin, M., Sotiropoulou, P. A. & Passegue, E. DNA-damage response in tissue-specific and cancer stem cells. *Cell Stem Cell* 8, 16–29 (2011).
- 114. Hovinga, K. E. et al. Inhibition of notch signaling in glioblastoma targets cancer stem cells via an endothelial cell intermediate. *Stem Cells* 28, 1019–1029 (2010).
- Chaudhary, P. M. & Roninson, I. B. Expression and activity of P-glycoprotein, a multidrug efflux pump, in human hematopoietic stem cells. *Cell* 66, 85–94 (1991).
- 116. Zhou, S. et al. The ABC transporter Bcrp1/ABCG2 is expressed in a wide variety of stem cells and is a molecular determinant of the side-population phenotype. Nat. Med. 7, 1028–1034 (2001).
- 117. Lin, T., Islam, O. & Heese, K. ABC transporters, neural stem cells and neurogenesis—a different perspective. *Cell Res.* 16, 857–871 (2006)
- Cell Res. 16, 857–871 (2006).

 118. Hirschmann-Jax, C. et al. A distinct "side population" of cells with high drug efflux capacity in human tumor cells. Proc. Natl Acad. Sci. USA 101, 14228–14233 (2004).
- 119. Stupp, R. et al. Effects of radiotherapy with concomitant and adjuvant temozolomide versus radiotherapy alone on survival in glioblastoma in a randomised phase III study: 5-year analysis of the EORTC-NCIC trial. *Lancet Oncol.* 10, 459–466 (2009).
- Singh, S. K. et al. Identification of a cancer stem cell in human brain tumors. *Cancer Res.* 63, 5821–5828 (2003).
- Bao, S. et al. Glioma stem cells promote radioresistance by preferential activation of the DNA damage response. *Nature* 444, 756–760 (2006).
- 122. Dent, P. et al. CHK1 inhibitors in combination chemotherapy: thinking beyond the cell cycle. *Mol. Interv.* 11, 133–140 (2011).
- 123. Ong, D. S. T. et al. PAF promotes stemness and radioresistance of glioma stem cells. *Proc. Natl Acad. Sci. USA* 114, E9086–E9095 (2017).
- 124. Druker, B. J. et al. Effects of a selective inhibitor of the Abl tyrosine kinase on the growth of Bcr-Abl positive cells. *Nat. Med.* 2, 561–566 (1996).
- 125. Kimura, S. Current status of ABL tyrosine kinase inhibitors stop studies for chronic myeloid leukemia. Stem Cell. Investig. 3, 36 (2016).
 126. Bhatia, R. et al. Persistence of malignant
- 126. Bhatia, R. et al. Persistence of malignant hematopoietic progenitors in chronic myelogenous leukemia patients in complete cytogenetic remission following imatinib mesylate treatment. *Blood* 101, 4701–4707 (2003).
- 127. Chomel, J. C. et al. Leukemic stem cell persistence in chronic myeloid leukemia patients with sustained undetectable molecular residual disease. *Blood* 118, 3657–3660 (2011).
- Graham, S. M. et al. Primitive, quiescent, Philadelphiapositive stem cells from patients with chronic myeloid leukemia are insensitive to STI571 in vitro. *Blood* 99, 319–325 (2002).

- 129. Corbin, A. S. et al. Human chronic myeloid leukemia stem cells are insensitive to imatinib despite inhibition of BCR-ABL activity. *J. Clin. Invest.* 121, 396–409 (2011).
- Jabbour, E. J., Cortes, J. E. & Kantarjian, H. M. Tyrosine kinase inhibition: a therapeutic target for the management of chronic-phase chronic myeloid leukemia *Expert Rev. Anticancer Ther.* 13, 1433–1452 (2013).
- 131. Chen, Y., Hu, Y., Zhang, H., Peng, C. & Li, S. Loss of the Alox5 gene impairs leukemia stem cells and prevents chronic myeloid leukemia. *Nat. Genet.* 41, 783–792 (2009).
- 132. Hu, Y., Chen, Y., Douglas, L. & Li, S. beta-Catenin is essential for survival of leukemic stem cells insensitive to kinase inhibition in mice with BCR-ABL-induced chronic myeloid leukemia. *Leukemia* 23, 109–116 (2009).
- 133. Zhao, C. et al. Hedgehog signalling is essential for maintenance of cancer stem cells in myeloid leukaemia. *Nature* 458, 776–779 (2009).
- 134. Shien, K. et al. Acquired resistance to EGFR inhibitors is associated with a manifestation of stem cell-like properties in cancer cells. *Cancer Res.* 73, 3051–3061 (2013).
- 135. Arasada, R. R., Amann, J. M., Rahman, M. A., Huppert, S. S. & Carbone, D. P. EGFR blockade enriches for lung cancer stem-like cells through Notch3-dependent signaling. *Cancer Res.* 74, 5572–5584 (2014).
- 136. Hu, S. et al. Antagonism of EGFR and Notch limits resistance to EGFR inhibitors and radiation by decreasing tumor-initiating cell frequency. Sci. Transl Med. 9, eaag0339 (2017).
- Brahmer, J. R. et al. Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. N. Engl. J. Med. 366, 2455–2465 (2012).
- 138. Iwai, Y. et al. Involvement of PD-L1 on tumor cells in the escape from host immune system and tumor immunotherapy by PD-L1 blockade. *Proc. Natl Acad. Sci. USA* 99, 12293–12297 (2002).
- 139. Leach, D. R., Krummel, M. F. & Allison, J. P. Enhancement of antitumor immunity by CTLA-4 blockade. *Science* **271**, 1734–1736 (1996).
- Ribas, A. & Wolchok, J. D. Cancer immunotherapy using checkpoint blockade. *Science* 359, 1350–1355 (2018).
- 141. Malta, T. M. et al. Machine learning identifies stemness features associated with oncogenic dedifferentiation. *Cell* **173**, 338–354 (2018).
- dedifferentiation. *Cell* **173**, 338–354 (2018). 142. Zaretsky, J. M. et al. Mutations associated with acquired resistance to PD-1 blockade in melanoma. *N. Engl. J. Med.* **375**, 819–829 (2016).
- Ji, R. R. et al. An immune-active tumor microenvironment favors clinical response to ipilimumab. Cancer Immunol. Immunother. 61, 1019–1031 (2012).
- 144. Spranger, S., Bao, R. & Gajewski, T. F. Melanoma-intrinsic β-catenin signalling prevents anti-tumour immunity. Nature 523, 231–235 (2015). This study demonstrates that stem cell signals within a tumour can lead to the exclusion of T cell infiltration and resistance to checkpoint inhibitor therapy, illustrating the impact of stem cell signalling on the microenvironment to promote
- aggressive disease.

 145. Cheah, M. T. et al. CD14-expressing cancer cells establish the inflammatory and proliferative tumor microenvironment in bladder cancer. *Proc. Natl Acad. Sci. USA* 112, 4725–4730 (2015).
- 146. Calabrese, C. et al. A perivascular niche for brain tumor stem cells. *Cancer Cell* **11**, 69–82 (2007).
- 147. Bao, S. et al. Stem cell-like glioma cells promote tumor angiogenesis through vascular endothelial growth factor. *Cancer Res.* 66, 7843–7848 (2006).
- 148. Folkins, C. et al. Glioma tumor stem-like cells promote tumor angiogenesis and vasculogenesis via vascular endothelial growth factor and stromal-derived factor 1. Cancer Res. 69, 7243–7251 (2009).
- 149. Charles, N. et al. Perivascular nitric oxide activates notch signaling and promotes stem-like character in PDGF-induced glioma cells. *Cell Stem Cell* 6, 141–152 (2010).
- Pietras, A. et al. Osteopontin-CD44 signaling in the glioma perivascular niche enhances cancer stem cell phenotypes and promotes aggressive tumor growth. Cell Stem Cell 14, 357–369 (2014).
- 151. Vredenburgh, J. J. et al. Phase II trial of bevacizumab and irinotecan in recurrent malignant glioma. *Clin. Cancer Res.* 13, 1253–1259 (2007).
- 152. Das, B. et al. Hypoxia enhances tumor stemness by increasing the invasive and tumorigenic side population fraction. Stem Cells 26, 1818–1830 (2008).
- 153. Heddleston, J. M., Li, Z., McLendon, R. E., Hjelmeland, A. B. & Rich, J. N. The hypoxic

- microenvironment maintains glioblastoma stem cells and promotes reprogramming towards a cancer stem cell phenotype. *Cell Cycle* **8**, 3274–3284 (2009).
- 154. Chiou, S. H. et al. BLIMP1 induces transient metastatic heterogeneity in pancreatic cancer. *Cancer Discov.* 7, 1184–1199 (2017).
- Discov. 7, 1184–1199 (2017).

 155. Tammela, T. et al. A Wnt-producing niche drives proliferative potential and progression in lung adenocarcinoma. Nature 545, 355–359 (2017).

 This study demonstrates how non-stem cells of the niche utilize stem cell signals to drive the tumorgenicity of neighbouring responder cells, illustrating the influence of the microenvironment on promoting aggressive disease through the use of stem cell signals.
- 156. Wang, X. et al. Reciprocal signaling between glioblastoma stem cells and differentiated tumor cells promotes malignant progression. *Cell Stem Cell* 22, 514–528 (2018).
- 157. Su, S. et al. CD10(+)GPR77(+) cancer-associated fibroblasts promote cancer formation and chemoresistance by sustaining cancer stemness. *Cell* 172, 841–856 (2018). In this study, cancer-associated fibroblasts are
 - In this study, cancer-associated fibroblasts are shown to support the CSC niche and contribute to therapy resistance, a key example of how the microenvironment impacts aggressive disease.
- 158. Chen, W. J. et al. Cancer-associated fibroblasts regulate the plasticity of lung cancer stemness via paracrine signalling. *Nat. Commun.* 5, 3472 (2014).
- 159. Sneddon, J. B. et al. Bone morphogenetic protein antagonist gremlin 1 is widely expressed by cancer-associated stromal cells and can promote tumor cell proliferation. Proc. Natl Acad. Sci. USA 103, 14842–14847 (2006).
- 160. Vermeulen, L. et al. Wnt activity defines colon cancer stem cells and is regulated by the microenvironment. Nat. Cell Biol. 12, 468–476 (2010).
- 161. Bajaj, J. et al. CD98-mediated adhesive signaling enables the establishment and propagation of acute myelogenous leukemia. *Cancer Cell* 30, 792–805 (2016).
- 162. Kwon, H. Y. et al. Tetraspanin 3 is required for the development and propagation of acute myelogenous leukemia. Cell Stem Cell 17, 152–164 (2015).
- 163. Passaro, D. et al. CXCR4 is required for leukemiainitiating cell activity in T cell acute lymphoblastic leukemia. *Cancer Cell* 27, 769–779 (2015).
 164. Ebinger, S. et al. Characterization of rare, dormant,
- 164. Ebinger, S. et al. Characterization of rare, dormant, and therapy-resistant cells in acute lymphoblastic leukemia. Cancer Cell 30, 849–862 (2016).
- 165. Sato, T. et al. Single Lgr5 stem cells build crypt-villus structures in vitro without a mesenchymal niche. *Nature* **459**, 262–265 (2009).
 - This is the first study to report the development of organoids that recapitulate the stem cell hierarchy of the original tissue.
- 166. Drost, J. & Clevers, H. Organoids in cancer research. Nat. Rev. Cancer 18, 407–418 (2018).
- 167. Roerink, S. F. et al. Intra-tumour diversification in colorectal cancer at the single-cell level. *Nature* 556, 457–462 (2018).
- 168. Boj, S. F. et al. Organoid models of human and mouse ductal pancreatic cancer. *Cell* 160, 324–338 (2015).
- 169. Tiriac, H. et al. Organoid profiling identifies common responders to chemotherapy in pancreatic cancer.

- Cancer Discov. https://doi.org/10.1158/2159-8290.CD-18-0349 (2018).
- Sachs, N. et al. A living biobank of breast cancer organoids captures disease heterogeneity. *Cell* 172, 373–386 (2018).
- Broutier, L. et al. Human primary liver cancer-derived organoid cultures for disease modeling and drug screening. Nat. Med. 23, 1424–1435 (2017).
- 172. Lee, S. H. et al. Tumor evolution and drug response in patient-derived organoid models of bladder cancer. Cell 173, 515–528 (2018).
- Cell 173, 515–528 (2018).

 173. Danial, C., Sarin, K. Y., Oro, A. E. & Chang, A. L. An investigator-initiated open-label trial of sonidegib in advanced basal cell carcinoma patients resistant to vismodegib. Clin. Cancer Res. 22, 1325–1329 (2016).
- 174. Sekulic, A. et al. Long-term safety and efficacy of vismodegib in patients with advanced basal cell carcinoma: final update of the pivotal ERIVANCE BCC study. BMC Cancer 17, 332 (2017).
- 175. Chang, A. L. et al. Safety and efficacy of vismodegib in patients with basal cell carcinoma nevus syndrome: pooled analysis of two trials. *Orphanet J. Rare Dis.* 11, 120 (2016).
- 176. Sekulic, A. et al. Efficacy and safety of vismodegib in advanced basal-cell carcinoma. N. Engl. J. Med. 366, 2171–2179 (2012).
- 177. Robinson, G. W. et al. Vismodegib exerts targeted efficacy against recurrent sonic hedgehog-subgroup medulloblastoma: results from phase II Pediatric Brain Tumor Consortium studies PBTC-025B and PBTC-032. J. Clin. Oncol. 33, 2646–2654 (2015).
- 178. Belani, C. P. et al. Vismodegib or cixutumumab in combination with standard chemotherapy for patients with extensive-stage small cell lung cancer: a trial of the ECOG-ACRIN Cancer Research Group (E1508). Cancer 122, 2371–2378 (2016).
- 179. US National Library of Medicine. ClinicalTrials.gov https://clinicaltrials.gov/ct2/show/NCT00833417 (2015).
- US National Library of Medicine. ClinicalTrials.gov https://clinicaltrials.gov/ct2/show/NCT01529450
- Taipale, J. et al. Effects of oncogenic mutations in Smoothened and Patched can be reversed by cyclopamine. *Nature* 406, 1005–1009 (2000).
- 182. Átwood, S. X., Li, M., Lee, A., Tang, J. Y. & Oro, A. E. GLI activation by atypical protein kinase C ι /λ regulates the growth of basal cell carcinomas. *Nature* 494, 484–488 (2013).
- 183. Rimkus, T. K., Carpenter, R. L., Qasem, S., Chan, M. & Lo, H. W. Targeting the sonic hedgehog signaling pathway: review of smoothened and GLI inhibitors. *Cancers* 8, E22 (2016).
- 184. US National Library of Medicine. ClinicalTrials.gov https://clinicaltrials.gov/ct2/show/NCT03035253 (2018).
- Lyou, Y., Habowski, A. N., Chen, G. T. & Waterman, M. L. Inhibition of nuclear Wnt signalling: challenges of an elusive target for cancer therapy. *Br. J. Pharmacol.* 174, 4589 –4599 (2017).
- 186. US National Library of Medicine. ClinicalTrials.gov https://clinicaltrials.gov/ct2/show/NCT01606579 (2017).
- US National Library of Medicine. ClinicalTrials.gov https://clinicaltrials.gov/ct2/show/NCT01345201 (2016).

- 188. Jimeno, A. et al. A first-in-human phase 1 study of the anti-cancer stem cell agent ipafricept (OMP-54F28), a decoy receptor for Wnt ligands, in patients with advanced solid tumors. Clin. Cancer Res. 23, 7490–7497 (2017).
- 189. US National Library of Medicine. ClinicalTrials.gov https://clinicaltrials.gov/ct2/show/NCT01351103 (2018).
- Jain, P. et al. Prognostic factors and survival outcomes in patients with chronic myeloid leukemia in blast phase in the tyrosine kinase inhibitor era: cohort study of 477 patients. Cancer 123, 4391–4402 (2017).
- of 477 patients. *Cancer* **123**, 4391–4402 (2017).
 191. Knoblich, J. A. Mechanisms of asymmetric stem cell division. *Cell* **132**, 583–597 (2008).
- 192. Knoblich, J. A. Asymmetric cell division: recent developments and their implications for tumour biology. *Nat. Rev. Mol. Cell Biol.* 11, 849–860 (2010).
 193. Bajaj, J., Zimdahl, B. & Reya, T. Fearful symmetry:
- 193. Bajaj, J., Zimdahl, B. & Reya, T. Fearful symmetry: subversion of asymmetric division in cancer development and progression. *Cancer Res.* 75, 792–797 (2015).
- 194. Ito, K. & Hotta, Y. Proliferation pattern of postembryonic neuroblasts in the brain of Drosophila melanogaster. *Dev. Biol.* 149, 134–148 (1992).
 195. Heidel, F. H. et al. The cell fate determinant Llgl1
- 195. Heidel, F. H. et al. The cell fate determinant Lig11 influences HSC fitness and prognosis in AML. J. Exp. Med. 210, 15–22 (2013).
- 196. He, L. et al. A microRNA component of the p53 tumour suppressor network. *Nature* 447, 1130–1134 (2007).
 197. Bu, P. et al. A microRNA miR-34a-regulated bimodal
- 197. Bu, P. et al. A microRNA miR-34a-regulated bimodal switch targets Notch in colon cancer stem cells. *Cell Stem Cell* 12, 602–615 (2013).
- 198. Li, Y. et al. MicroRNA-34a inhibits glioblastoma growth by targeting multiple oncogenes. *Cancer Res.* 69, 7569–7576 (2009).
- 199. Hwang, W. L. et al. MicroRNA-146a directs the symmetric division of Snail-dominant colorectal cancer stem cells. Nat. Cell Biol. 16, 268–280 (2014).
- 200. Chen, G. et al. Human Brat ortholog TRIM3 is a tumor suppressor that regulates asymmetric cell division in glioblastoma. *Cancer Res.* 74, 4536–4548 (2014).
- Wang, L. et al. A long non-coding RNA targets microRNA miR-34a to regulate colon cancer stem cell asymmetric division. *eLife* 5, e14620 (2016).

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Author contributions

N.K.L., A.G.B. and T.R. researched data for the article, wrote the article and reviewed or edited the manuscript before submission. N.K.L. and T.R. also made substantial contributions to the discussion of content.

Competing interests

The authors declare no competing interests.

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